

Microbial strategies for antagonizing Toll-like-receptor signal transduction

Charles V Rosadini and Jonathan C Kagan



Within a few years of the discovery of Toll-like receptors (TLRs) and their role in innate immunity, viral and bacterial proteins were recognized to antagonize TLR signal transduction. Since then, as TLR signaling networks were unraveled, microbial systems have been discovered that target nearly every component within these pathways. However, recent findings as well as some notable exceptions promote the idea that more of these systems have yet to be discovered. For example, we know very little about microbial systems for directly targeting non-cytoplasmic portions of TLR signaling pathways, that is, the ligand interacting portions of the receptor itself. In this review, we compare and contrast strategies by which bacteria and viruses antagonize TLR signaling networks to identify potential areas for future research.

Addresses

Harvard Medical School and Division of Gastroenterology, Boston Children's Hospital, Boston, MA 02115, USA

Corresponding author: Kagan, Jonathan C
(jonathan.kagan@childrens.harvard.edu)

Current Opinion in Immunology 2015, **32**:61–70

This review comes from a themed issue on **Innate immunity**

Edited by **Zhijian J Chen** and **Sebastian Amigorena**

For a complete overview see the [Issue](#) and the [Editorial](#)

Available online 20th January 2015

<http://dx.doi.org/10.1016/j.coi.2014.12.011>

0952-7915/© 2015 Elsevier Ltd. All rights reserved.

Introduction

Innate immunity is mediated by specialized proteins called pattern recognition receptors that sense microbial invaders and guide our immune systems to eradicate infections. These receptors detect pathogen associated molecular patterns (PAMPs), which are structures common to many microbial species, such as viral nucleic acids or bacterial lipopolysaccharides (LPS). The first identified and most studied group of these receptors, Toll-like-receptors (TLRs), are displayed at the cell surface and within endosomal compartments where they act as molecular sentinels to detect invading microbes. Upon recognition of a pathogen, in either infected or non-infected cells, TLRs initiate a signaling cascade that leads to expression and release of pro-inflammatory cytokines, chemokines, and Type-I interferons.

Cytokine and interferon expression leads to recruitment and activation of immune cells to promote clearance of the infectious agent, but also stimulates expression of genes to block entry and/or replication of microbes. This elegant system is adapted to detect and eliminate almost any threat. However, pathogenic microbes are masters of evading host innate immunity and have evolved a multitude of mechanisms for preventing the antimicrobial activities of TLR signaling pathways. Microbial manipulation of host TLR signaling comes in the form of three main strategies: Firstly, directly antagonize signaling components; secondly, avoiding detection by altering their PAMPs to be less immunogenic and finally, disrupting cellular organelles/induce cell death. Here we will focus our discussion on direct antagonism approaches.

An abundance of viral proteins are known to antagonize TLR signaling at almost every signaling step (reviewed [1]), from blocking signaling intermediates to inhibition of downstream transcription factor activation. While much less is understood about how bacteria block TLR pathways, recent work in this area suggests that they too antagonize several steps of these pathways. We postulated that by surveying the known systems utilized by viruses and bacteria to block TLR signaling, patterns would emerge that would allow us to predict where future research might be most productive. Microbial strategies were organized by the step, or the module, of TLR receptor pathways that they antagonize (**Table 1**). The receptor module (**Figure 1**) consists of binding of ligands by various TLRs (reviewed [2]), receptor dimerization, and recruitment of the receptor to subcellular signaling sites such as phosphatidylinositol 4,5-bisphosphate (PIP2) rich regions of the plasma membrane, or phosphatidylinositol 3-phosphate (PI3P) rich endosomes [3,4]. Once the receptor has moved to signaling sites, it can associate with intracellular sorting adaptor proteins, which by virtue of their association with lipids such as PIP2 and PI3P, are prepositioned on specific organelles to detect activated TLRs. Sorting adaptors, TIRAP and TRAM, promote recruitment of signaling adaptors, MyD88 and TRIF, respectively [5], which are thought to trigger the formation of higher order filamentous structures called Supramolecular Organizing Centers (SMOC) (**Figure 1**) [6]. SMOCs are multiprotein complexes that have been proposed to serve as organizing centers that coordinate the multitude of cellular responses to microbial infections and cytokines [6]. The best-characterized SMOC is that formed between

Table 1

Microbial antagonization of TLR signaling modules.

Module targeted	Microbe	Protein	Mode of antagonization	Citation	
Receptor	<i>Staphylococcus aureus</i>	SSL3, SSL4	Bind TLR2 ectodomain	[64**,65**]	
SMOC	African swine fever virus	ORF I329L	Possibly targets TRIF	[70]	
	Coxsackievirus	3C	Cleaves TRIF	[11]	
	Enterovirus 68	3C	Cleaves TRIF	[71]	
	Hepatitis C virus (HCV)	NS3-4A	Cleaves TRIF	[72]	
	HCV	NS5A	Binds MyD88	[73]	
	HSV-1	ICP0	Promotes degradation of Myd88 and TIRAP	[17]	
	KSHV	RTA	Degrades TRIF	[74]	
	Vaccinia virus (VACV)	A46R	Binds TIRAP, TRAM, MyD88, TRIF, TLR4	[14,15]	
	VACV	A52R	Binds IRAK2	[20]	
	<i>Brucella</i> spp.	BtpB	Probable TIR domain containing	[75]	
	<i>Brucella melitensis</i>	TcpB	Blocks TIR–TIR interactions and promotes degradation of MyD88 and TIRAP	[76]	
	<i>Escherichia coli</i>	TcpC	Blocks TIR–TIR interactions	[76]	
	<i>S. aureus</i>	TirS	Blocks TIR–TIR interactions	[13]	
	<i>Salmonella enterica</i> serovar <i>Enteritidis</i>	<i>TlpA</i>	Blocks TIR–TIR interactions	[56]	
	Yersinia <i>pestis</i>	YpTdp		Blocks TIR–TIR interactions	[12]
TRAF6/TAK1		Herpes simplex virus (HSV-1)	Us3	Decreases levels of TRAF6 polyubiquitination	[22]
		Epstein-Barr virus (EBV)	BPLF1	Deubiquitinates TRAF6	[23,24*]
		<i>Shigella flexneri</i>	OspI	Deamidates UBC13	[25**]
		Enteropathogenic <i>E. coli</i> (EPEC)	NleE	Modifies TAB2 and TAB3	[26**]
	<i>Yersinia</i> spp.	YopJ	Acetylates TAK1	[27**]	
MAP kinases	Ebola virus	VP24	Blocks phosphorylation of p38	[29]	
	Hepatitis B virus	HBsAg	Inhibits phosphorylation of JNK1/2 and c-Jun	[30*]	
	<i>Bacillus anthracis</i>	LF	Degrades MAPKK 1 and 2	[77]	
	EPEC	NleC and NleD	Cleaves JNK	[51]	
	<i>Salmonella typhimurium</i>	AvrA	Acetylates MKK4	[78]	
	<i>Salmonella</i> spp.	SpvC	Modifies c-Jun, Erk1/2, and p38	[57,58]	
	<i>S. flexneri</i>	OspF	Modifies c-Jun, Erk1/2, and p38	[57,79]	
	<i>Vibrio parahemolyticus</i>	VopA	Acetylates MAPKK	[80]	
	<i>Yersinia</i> spp.	YopJ	Acetylates MKK6, MKK4, MKK7	[27**,59]	
	NEMO complex	EBV	BPLF1	Deubiquitinates NEMO	[24*]
Foot-and-mouth disease virus		3C	Cleaves NEMO	[32]	
Hepatitis A virus		3C	Cleaves NEMO	[33]	
HCV		NS3	Binds TBK1	[81]	
HSV-1		γ 34.5	Binds TBK1	[82]	
Mouse hepatitis virus A59		NSp3 (PLP2 domain)	Deubiquitinates TBK1	[83]	
Sendai virus		C-protein	Binds IKK α	[34]	
VACV		B14R	Binds IKK β	[84]	
VACV		C6	Binds TANK, SINTBAD, or NAP1	[85]	
VACV		N1L	Associates with IKK complex and TBK1	[55]	
VACV		K7R	Binds DDX3	[86]	
<i>S. flexneri</i>		IpaH9.8	Promotes NEMO degradation	[87]	
<i>Yersinia</i> spp.		YopJ/P	Acetylates IKK β	[88]	
Transcription factors		Classical Swine Fever Virus	Npro	Promotes IRF3 degradation	[89]
		EBV	BPLF1	Deubiquitinates I κ B α	[24*]
	HIV	Vpr, Vif	Degrade IRF3	[90,91]	
	HIV	Vpu	Possibly blocks IRF3 and NF- κ B	[91,92]	
	HSV-1	ICP27	Stabilizes I κ B α	[37]	
	HSV-1	ICP0	Sequesters IRF3–CBP/p300	[43]	
	HSV-1	Us3	Phosphorylates IRF3 and NF- κ B	[41,42]	
	HSV-1	VP16	Binds CBP in the nucleus	[45]	
	KSHV	RTA	Promotes degradation of IRF3 and IRF7	[93]	
	KSHV	K-bZIP	Competes for IRF3 binding sites	[94]	

Table 1 (Continued)

Module targeted	Microbe	Protein	Mode of antagonization	Citation
	KSHV	vIRF-1	Inhibits IRF3 interaction with CBP and p300	[44]
	KSHV	vIRF3	Binds IRF3	[95]
	Measles virus	C-protein	Unknown function in nucleus	[96]
	Measles virus	V-protein	Binds NF- κ B and IRF3	[97]
	Mumps virus	V-protein	Mimics IR3	[98]
	Rotavirus	NSP1	Degrades IRF3, IRF7 or E3 ligase β -TrCP	[38,99,100]
	Sendai virus and New castle disease virus	V-protein	Binds active IRF3 and prevents nuclear translocation	[97]
	VACV	A49	Binds and inhibits β -TrCP	[39]
	Varicella-Zoster virus (VZV)	ORF47	Atypically phosphorylates IRF3	[40]
	VZV	ORF61	Ubiquitinates IRF3 and NF- κ B	[101,102]
	EPEC	NleC,NleD	Cleaves NF- κ B	[48–51]
	EPEC	NleB, NleE	Inhibits nuclear translocation of NF- κ B	[52]
	EPEC and enterohemorrhagic <i>Escherichia coli</i> (EHEC)	NleH1, NleH2	Inhibits nuclear translocation of NF- κ B	[46,47]
	<i>S. flexneri</i>	OspZ	Inhibits nuclear translocation of NF- κ B	[52]
	<i>S. flexneri</i>	OspG	Binds and interferes with UbcH5b	[53]

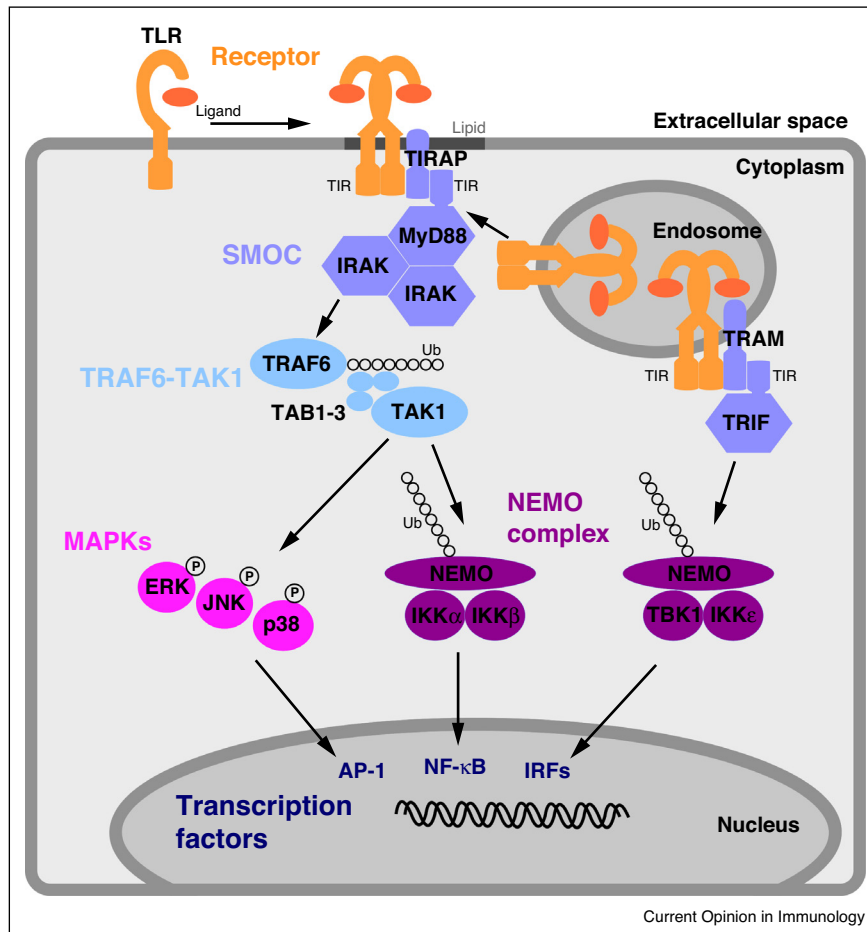
the aforementioned TIRAP and MyD88 adaptors and IRAK kinases. This SMOc has been dubbed the myddosome [4,7,8]. Myddosome formation activates a signaling module consisting of an E3 ubiquitin ligase called TRAF6 and the TAK1 complex (Figure 1). TAK1 phosphorylates proteins leading to activation of two signaling modules: the mitogen activated protein kinases (MAPKs) and the NEMO complex (Figure 1). Signal transduction through MAPKs or the NEMO complex results in the final signaling module: activation and nuclear translocation of transcription factors (Figure 1), such as AP-1 and nuclear factor κ B (NF- κ B), respectively. In addition, the NEMO complex can also activate transcription factors of the interferon regulatory factor (IRF) family (reviewed [2,9]). Below we will describe recent findings of microbial systems to antagonize each TLR signaling module.

The SMOc

SMOc formation immediately follows receptor activation and is initiated by interaction between TLRs and sorting adaptor proteins found on specific organelles within the cell. This interaction occurs between the Toll/IL-1 receptor/resistance (TIR) domains present on both TLRs and adaptor proteins [10]. Sorting adaptors recruit signaling adaptors to promote formation of SMOcs, such as the myddosome [4,5]. The importance of the TLR-induced SMOcs in signal transduction can be revealed by the various pathogenic proteins that have been proposed to counteract the functions of the TIR domain containing adaptors. Microbial systems for antagonizing this signaling module mostly target adaptor proteins and are distributed fairly equally between viruses and bacteria. However, viruses and bacteria tend to rely on their own unique mechanisms of antagonism. For example, viruses heavily favor degrading these adaptor proteins, such as the 3C protease of coxsackievirus B that cleaves TRIF [11], where bacteria often use molecular mimicry by producing proteins that interfere with adaptor protein aggregation. Briefly, bacteria use elaborate secretion

systems to inject proteins called effectors into host cells. Some of these effectors contain TIR domains that interact with TIR domains of MyD88, TIRAP, TRIF, TRAM as well as the TLRs. TIR domain containing proteins of bacteria have been reviewed recently [10], however new findings of these proteins in *Yersinia pestis* and *Staphylococcus aureus* demonstrate that this field is rapidly growing and likely more TIR domain containing proteins that can block TLR signaling will emerge [12,13]. Interestingly, the first protein found to interfere with adaptors to inhibit TLR signaling, A46R of Vaccinia virus (VACV), functions similar to the bacterial TIR domain-containing proteins. A46R binds to MyD88, TIRAP, TRIF, TRAM and TLR4, and can prevent signal transduction [14,15]. This, however, is not the only example of overlap between microbial strategies for blocking adaptor proteins. In fact, an effector called TcpB from *Brucella* spp. binds TIRAP and induces its ubiquitination and subsequent degradation [16]. This mechanism is reminiscent of the actions of herpes simplex virus (HSV-1) immediate early protein ICP0 that targets MyD88 and TIRAP for degradation [17]. Interestingly, the degree of mimicry of TcpB for TIRAP is remarkable, in that (like TIRAP) it contains a TIR domain and a domain that binds to the plasma membrane via interactions with PIP2. Before inducing TIRAP degradation, TcpB binds to TIRAP and prevents its interaction with TLR4 [18,19]. Therefore, TcpB represents a unique example where a single protein binds and physically blocks but also promotes degradation of adaptors. Unlike ICP0, which potentially targets TIRAP via its E3 ubiquitin ligase activity, it remains unclear how TcpB induces TIRAP degradation. Future study of the function of this protein and its interactions with TIRAP will lead to greater understanding of the earliest modules of TLR signaling. Overall, findings of bacterial proteins that target the adaptors will likely continue. Intriguingly, while several microbial proteins engage myddosome components through TIR–TIR interactions, only one protein is known to antagonize kinases within the myddosome.

Figure 1



Modules of TLR signaling. *The receptor module*: binding of ligands to TLRs at the plasma membrane or within endosomes, receptor dimerization and migration to subcellular signaling sites including specific lipid rich regions within the membrane. *Supramolecular Organizing Center (SMOC)*: TLRs interact with intracellular sorting adaptor proteins, TIRAP and TRAM, which recruit signaling adaptors MyD88 and TRIF, respectively. Adaptor proteins and TLRs interact via TIR domains. Signaling adaptors trigger formation of SMOCs such as the myddosome which consists of TIRAP, MyD88, and IRAK kinases. *TRAF6 and TAK1 complex*: Myddosome formation activates the E3 ubiquitin (Ub) ligase TRAF6 which ubiquitinates itself and recruits TAB proteins. TAB proteins activate the MAP3K, TAK1. *Mitogen activated protein kinase (MAPK)*: TAK1 phosphorylates proteins of the MAPK cascade resulting in phosphorylation and activation of MAPKs, ERK, JNK and p38. *NEMO complex*: Ubiquitinated NEMO is required for IKK α /IKK β to be activated by TAK1 downstream of MyD88 signaling, but also to promote activation of TBK1 and IKK ϵ downstream of signaling through TRIF. *Transcription factors*: Transcription factor activation involves activation and nuclear translocation of AP-1 (through MAPKs), NF- κ B, through IKK α /IKK β , and the IRFs through TBK1, IKK ϵ , and in certain circumstances IKK α . Once in the nucleus transcription factors induce expression of proinflammatory cytokines, chemokines, and interferons.

The A52R protein of VACV binds IRAK2, a myddosome component, to inhibit signaling [20]. Future research will be needed to determine if other microbes utilize similar strategies to inhibit TLR signaling at the module of SMOC assembly or function.

TRAF6 and TAK1 complex

Myddosome formation involves the recruitment of IRAK family kinases, which in turn activate a module consisting of the E3 ubiquitin ligase TRAF6 and the MAP3K, TAK1. TRAF6 polyubiquitinates itself and recruits TAK1 regulatory components called TAK1-binding protein (TAB) 1–3. Binding of TAB2 and TAB3 to ubiquitin

chains made by TRAF6 is required to activate TAK1 [21]. Active TAK1 phosphorylates proteins of the MAPK cascade and also the NEMO complex, leading to their activation (discussed in detail below). In the last few years, several discoveries of both viral and bacterial mechanisms for antagonizing the TRAF6 and TAK1 complex have emerged. First, the kinase U_S3 of HSV-1 was shown to reduce levels of polyubiquitination on TRAF6, resulting in inhibition of TLR2 signaling during viral infection [22]. Inhibition of TLR2 signaling by U_S3 is dependent on its kinase activity and found to be downstream of MyD88. However, how this protein affects TRAF6 ubiquitination remains an open question

[22]. Next, recent work on Epstein–Barr virus protein BPLF1 suggests that this protein is a deubiquitinase that also inhibits ubiquitination of TRAF6 [23,24*]. Furthermore, OspI of *Shigella flexneri* is an example of a bacterial protein that inhibits TRAF6 polyubiquitination [25**]. However, this enzyme uses a unique approach by deamidating a glutamine residue in the E2 ubiquitin conjugating enzyme UBC13, which is required for ubiquitination of TRAF6 [25**]. Enzymes that target activation of the TAK1–TAB1–3 complex downstream of TRAF6 were also described, including NleE from Enteropathogenic *Escherichia coli* (EPEC) and YopJ from *Y. pestis* [26**,27**]. NleE is a methyltransferase that targets critical zinc coordinating cysteine residues within zinc-finger domains of TAB2 and TAB3. This novel mechanism results in loss of ubiquitin binding activity of TAB2 and TAB3, preventing activation of TAK1 and subsequently, NF- κ B [26**]. In contrast, YopJ inhibits TAK1 kinase activity by acetylating key residues in its active site [27**]. In this way YopJ blocks TAK1-dependent activation of MAP2Ks and inhibitor of NF- κ B kinases (IKKs). Interestingly, unlike what has been found for targeting many of the other modules of TLR signaling, the mechanisms for targeting this module are diverse and dissimilar between viruses and bacteria. Perhaps future research will uncover how common each of these mechanisms are across a broad range of pathogens. On the other hand, with so few examples described, potentially other fascinating mechanisms for blocking this module of signaling will be discovered.

MAPK cascade

The MAPK cascade is a phospho-relay system that signals through ERK, JNK and p38 to activate the transcription factor AP-1. Previously, only bacterial systems were known to block MAPK cascades (reviewed [28]). However, recent research revealed viral proteins capable of targeting MAPKs, including VP24 of Ebola virus, which inhibits interferon- β (IFN- β) stimulated JAK-STAT signaling by blocking phosphorylation of p38 [29]. Furthermore, the surface antigen of Hepatitis B virus (HBsAg) blocks TLR2 signaling by inhibiting phosphorylation of JNK-1/2 and c-Jun, although the mechanism for this inhibition remains unclear [30*]. Together these examples not only demonstrate that viruses have the ability to block TLR signaling at this module, but also suggest that viruses target select MAPKs. The advantage of targeting select MAPKs may be to ensure maintenance of other MAPK-dependent pathways that promote survival of the infected cell.

NEMO complex

Downstream of MyD88 and TAK1 is a signaling module governed by the protein NF- κ B essential modulator (NEMO). NEMO controls the activation of IKKs, such as IKK α and IKK β . NEMO binds non-degradative polyubiquitin chains that act as a scaffold for TAK1

phosphorylation of IKK α /IKK β [21]. IKK α /IKK β , in turn phosphorylates the inhibitor of NF- κ B, I κ B, promoting its ubiquitination and subsequent degradation, releasing NF- κ B so it can translocate to the nucleus. In the case of endosomal TLRs, IKK α can also promote IRF7 activation. Alternatively, signaling downstream of TRIF leads to NEMO complex activation, however this results in recruitment and activation of TBK1/IKK ϵ and their accessories TANK, SINTBAD, and NAP1. These proteins function to activate the transcription factor IRF3, which is a master regulator of interferon expression [31].

Diverse microbial strategies are used to target the NEMO complex including degrading, binding, and even deubiquitinating host enzymes and accessory proteins. Recent discoveries revealed that the 3C proteases from hepatitis A virus and foot-and-mouth disease virus can inhibit TLR signaling by cleaving NEMO [32,33]. Additionally, C-protein of Sendai virus blocks TLR7 and TLR9 signaling by binding IKK α and inhibiting phosphorylation of IRF7 [34]. Furthermore, BPLF1 of Epstein–Barr virus blocks signaling by deubiquitinating IKK α and NEMO [24*]. Interestingly, although a limited number of bacterial proteins are known to target this module, one such effector, IpaH9.8 of *S. flexneri* utilizes a strategy similar to viruses by promoting the degradation of NEMO. This example of bacteria using a strategy similar to that of viruses promotes the idea that knowledge of viral strategies can be used as a framework for future investigation of novel bacterial proteins that antagonize this signaling module.

Transcription factors

Transcription factor activation involves nuclear translocation of AP-1, NF- κ B and the IRFs, which lead to transcriptional responses and the production of pro-inflammatory cytokines, chemokines, and interferons. Targeting the transcription factor module is a widely used strategy for viruses. Viral mechanisms for targeting this module include degrading and mimicking transcription factors and have been extensively reviewed elsewhere [35,36]; therefore we will focus on a few recent findings that highlight some of their tactics. First, HSV-1 and rotavirus prevent NF- κ B translocation into the nucleus by stabilizing or preventing the degradation of I κ B [37,38]. This tactic is also used by VACV, whose protein A49 inhibits this signaling module by binding and inhibiting the activity of β -TrCP, an E3 ligase required for ubiquitination and degradation of I κ B [39]. A second mechanism is that used by ORF47 of varicella-zoster virus and U $_S$ 3 of HSV-1, which phosphorylate IRF3 to block its proper activation by host kinases [40,41]. Recently, U $_S$ 3 was also shown to hyperphosphorylate NF- κ B, blocking its activation [42]. Thirdly, some viruses bind co-activators such as CBP and p300 in the nucleus, an activity that prevents transcription factors from reaching their target genes. This method is used by herpesvirus

proteins including ICP0 of HSV-1 and vIRF-1 of Kaposi's sarcoma-associated herpesvirus (KSHV) [43,44]. Recent data suggest that VP16 of HSV-1 is also capable of binding to CBP in the nucleus [45].

Despite the abundance of viral species that block this module, only a few bacterial species are known to possess this ability. These include enterohemorrhagic *E. coli* (EHEC), EPEC, and *S. flexneri*. EHEC and EPEC encode effectors NleH1 and NleH2 that block NF- κ B by binding to NF- κ B subunit, RPS3 [46,47]. NleH1 but not NleH2 was shown to inhibit nuclear translocation of RPS3, however both NleH1 and NleH2 have also been implicated in preventing ubiquitination and degradation of I κ B [47]. EPEC also encodes NleC and NleD which are zinc metalloproteases that cleave the RelA subunit of NF- κ B to prevent its activation [48–51]. Two other EPEC proteins, NleB and NleE, also block NF- κ B translocation [52]. The targets of NleB and NleE are not known, however their mechanisms are likely different as NleB blocks signaling in response to stimulation of cells with TNF α but not IL-1 β , whereas, NleE blocks both TNF α and IL-1 β stimulated responses. *S. flexneri* encodes OspZ, which is interchangeable with the C-terminus of NleE, and thus likely acts via a similar mechanism [52]. A second *S. flexneri* encoded effector, OspG, acts similar to VACV A49 discussed above. OspG prevents degradation of I κ B by binding and interfering with the E2 ubiquitin-conjugating enzyme, UbcH5b [53,54]. This example of overlap between bacterial and viral tactics reinforces the idea that future investigation into bacterial proteins that antagonize TLR signaling can be guided by our vast knowledge of viral antagonization mechanisms.

Targeting multiple modules

A common tactic utilized by viruses and bacteria is to attack several modules of TLR signaling simultaneously. For example, VACV expresses N1L to block IKKs but also expresses A46R and A52R to target adaptor proteins and IRAK kinases [14,20,55], respectively, whereas *S. flexneri* expresses OspF which blocks MAPKs, OspI which blocks TRAF6, and IpaH9.8 which promotes degradation of NEMO. Other examples include *Salmonella* spp. which express TlpA, a TIR domain containing protein that interferes with adaptor proteins [56], and SpvC, a phosphothreonine lyase that irreversibly modifies MAP kinases [57,58]. It is also common for viruses and bacteria to utilize a single protein for blocking more than one module of TLR signaling. Examples include: BPLF1 of Epstein–Barr virus which deubiquitinates TRAF6, IKK α , and NEMO [23,24^{*}], ICP0 of HSV-1 which promotes degradation of TIRAP and MyD88 but also binds and sequesters IRF3 in the nucleus [17,43], and YopJ of *Yersinia* spp., which acetylates several MAPKs and IKK β [27^{**},59], effectively eliminating MyD88-dependent signaling through both the MAPK cascade and the NEMO complex. This strategy is not surprising for viruses as they

express a limited number of proteins and thus often utilize a single protein to perform diverse functions. It is more intriguing however for bacteria, which can inject dozens of proteins into host cells [60]. However, for some species that inject only a small number of proteins into host cells, such as *Y. pestis* which only secretes six known effectors [61], this strategy suggests a need for fine tuning of their existing proteins to target multiple host factors. Overall, the popularity of these strategies suggests that if a virus or bacterium is known to antagonize TLR signaling at one module, it will likely be found to target other modules.

Receptors

In our search for microbial systems that antagonize specific components of TLR signaling networks, we find that viruses are not known to antagonize TLR signaling at the module of the receptor. Perhaps this is due to a lack of effective tools, such as antibodies, for detecting changes in TLRs on the surface of cells during infections. We also note that this module of TLR signaling is the least understood. Assays to monitor inducible (microbe-triggered) interactions between TLRs and their ligands or co-receptors are very much lacking. Despite these technical challenges, it is possible that viruses may not interfere with TLR detection directly. One reason to consider this possibility is that it may be more efficient for them to target downstream signaling components that are used by multiple innate immune pathways than to antagonize individual receptors. Nevertheless, there are some viral strategies that could antagonize TLRs. For example, viral proteins in both human T-cell leukemia virus type 1 (p30) and HSV-1 (U $_s$ 3) can promote down-regulation of TLR4 and TLR3, respectively [62,63]. It remains unclear how common this strategy is for other viruses, and whether the down-regulation observed is sufficient to functionally inactivate the signaling pathway.

Despite the challenges of investigating the receptor module, recent discoveries have revealed that at least two bacterial proteins are capable of binding to TLRs and blocking their function. The secreted staphylococcal superantigen-like proteins (SSLs) SSL3 and SSL4 from *S. aureus* have the ability to interact with the ectodomain of TLR2 [64^{**},65^{**}]. Incubation of mouse or human leukocytes with purified SSL3 blocks TLR2-dependent responses to purified TLR2 ligands as well as heat killed *S. aureus*. However, the ability of *S. aureus* to suppress TLR2 responses in a SSL3/4 dependent manner during *S. aureus* infection has not been demonstrated. Thus, the relevance of this system for virulence and survival of *S. aureus* remains an open question. Interestingly, other SSL proteins of *S. aureus* have been shown to bind and block extracellular immune components on leukocytes such as SSL5 which binds chemokine and anaphylatoxin receptors [66]. Thus, SSLs demonstrate the potential for bacterial proteins to antagonize host immune functions by binding to extracellular components.

Other bacterial strategies also suggest possible mechanisms for direct TLR antagonism, although these strategies have never been directly implicated in antagonizing these receptors. For example, bacteria can secrete proteins to degrade host immune components such as the IgA proteases of *Streptococcus pneumoniae*, *Haemophilus influenzae*, and *Neisseria* spp., [67,68] and also secrete many other proteases that have been linked with virulence and survival of different bacterial species. Interestingly, the cysteine proteases, known as gingipains, from *Porphyromonas gingivalis* have been shown to preferentially cleave CD14, an extracellular co-receptor vital to TLR4 signaling, from phagocytes [69], though the relevance of this to survival and virulence of *Porphyromonas* spp. remains unclear. This example suggests that bacterial proteases have the potential to counteract TLR function and inflammatory responses through direct cleavage of receptor complexes on the surface of host cells. Perhaps future investigation of viral or bacterial proteins and their interaction with TLRs will reveal novel systems for targeting the receptor module.

Perspective

Recent years have seen an explosion in findings of bacterial and viral systems to antagonize TLR signaling. Here we have compiled a wide-ranging list of these known strategies with the intent of providing insight into where future research will be best applied. Notably, we have identified gaps in our knowledge, such as the lack of systems for targeting the receptor module, which present some interesting challenges for the field. Overall, our findings indicate that viruses and bacteria have evolved similar mechanisms for antagonizing TLR signaling. This evolutionary convergence of strategies suggests that the information presented here can be used to guide future discovery of novel microbial factors that antagonize TLR signaling, both viral and bacterial.

Acknowledgements

We would like to thank the members of the Kagan lab for helpful discussions. JCK is supported by NIH grants AI093589, AI072955 and P30 DK34854, and an unrestricted gift from Mead Johnson & Company. Dr. Kagan holds an Investigators in the Pathogenesis of Infectious Disease Award from the Burroughs Wellcome Fund. CVR is supported by NIH grant DK102317-01.

References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- of outstanding interest

1. Bowie AG, Unterholzner L: **Viral evasion and subversion of pattern-recognition receptor signalling.** *Nat Rev Immunol* 2008, **8**:911-922.
2. Kawai T, Akira S: **The role of pattern-recognition receptors in innate immunity: update on Toll-like receptors.** *Nat Immunol* 2010, **11**:373-384.
3. Kagan JC, Medzhitov R: **Phosphoinositide-mediated adaptor recruitment controls Toll-like receptor signaling.** *Cell* 2006, **125**:943-955.
4. Bonham KS, Orzalli MH, Hayashi K, Wolf AJ, Glanemann C, Weninger W, Iwasaki A, Knipe DM, Kagan JC: **A promiscuous lipid-binding protein diversifies the subcellular sites of Toll-like receptor signal transduction.** *Cell* 2014, **156**:705-716.
5. Kagan JC: **Signaling organelles of the innate immune system.** *Cell* 2012, **151**:1168-1178.
6. Kagan JC, Magupalli VG, Wu H: **SMOCs: supramolecular organizing centres that control innate immunity.** *Nat Rev Immunol* 2014, **14**:821-826.
7. Motshwene PG, Moncrieffe MC, Grossmann JG, Kao C, Ayaluru M, Sandercock AM, Robinson CV, Latz E, Gay NJ: **An oligomeric signaling platform formed by the Toll-like receptor signal transducers MyD88 and IRAK-4.** *J Biol Chem* 2009, **284**:25404-25411.
8. Lin SC, Lo YC, Wu H: **Helical assembly in the MyD88-IRAK4-IRAK2 complex in TLR/IL-1R signalling.** *Nature* 2010, **465**:885-890.
9. Cervantes JL, Weinerman B, Basole C, Salazar JC: **TLR8: the forgotten relative revindicated.** *Cell Mol Immunol* 2012, **9**:434-438.
10. Patterson NJ, Werling D: **To con protection: TIR-domain containing proteins (TcP) and innate immune evasion.** *Vet Immunol Immunopathol* 2013, **155**:147-154.
11. Mukherjee A, Morosky SA, Delorme-Axford E, Dybdahl-Sissoko N, Oberste MS, Wang T, Coyne CB: **The coxsackievirus B 3C protease cleaves MAVS and TRIF to attenuate host type I interferon and apoptotic signaling.** *PLoS Pathog* 2011, **7**:e1001311.
12. Rana RR, Simpson P, Zhang M, Jennions M, Ukegbu C, Spear AM, Alguel Y, Matthews SJ, Atkins HS, Byrne B: **Yersinia pestis TIR-domain protein forms dimers that interact with the human adaptor protein MyD88.** *Microb Pathog* 2011, **51**:89-95.
13. Askarian F, van Sorge NM, Sangvik M, Beasley FC, Henriksen JR, Sollid JU, van Strijp JA, Nizet V, Johannessen M: **A Staphylococcus aureus TIR domain protein virulence factor blocks TLR2-mediated NF-kappaB signaling.** *J Innate Immun* 2014, **6**:485-498.
14. Bowie A, Kiss-Toth E, Symons JA, Smith GL, Dower SK, O'Neill LA: **A46R and A52R from vaccinia virus are antagonists of host IL-1 and Toll-like receptor signaling.** *Proc Natl Acad Sci U S A* 2000, **97**:10162-10167.
15. Stack J, Haga IR, Schroder M, Bartlett NW, Maloney G, Reading PC, Fitzgerald KA, Smith GL, Bowie AG: **Vaccinia virus protein A46R targets multiple Toll-like-interleukin-1 receptor adaptors and contributes to virulence.** *J Exp Med* 2005, **201**:1007-1018.
16. Sengupta D, Koblansky A, Gaines J, Brown T, West AP, Zhang D, Nishikawa T, Park SG, Roop RM 2nd, Ghosh S: **Subversion of innate immune responses by Brucella through the targeted degradation of the TLR signaling adapter, MAL.** *J Immunol* 2010, **184**:956-964.
- Ectopic expression of TcpB in 293 cells reduces TLR4 signaling, TIRAP phosphorylation, and levels of TIRAP protein. TcpB expression promoted enhanced polyubiquitination of TIRAP. Finally, TIRAP levels were reduced during *B. abortus* infections of J774 cells in a TcpB specific manner.
17. van Lint AL, Murawski MR, Goodbody RE, Severa M, Fitzgerald KA, Finberg RW, Knipe DM, Kurt-Jones EA: **Herpes simplex virus immediate-early ICP0 protein inhibits Toll-like receptor 2-dependent inflammatory responses and NF-kappaB signaling.** *J Virol* 2010, **84**:10802-10811.
18. Radhakrishnan GK, Yu Q, Harms JS, Splitter GA: **Brucella TIR domain-containing protein mimics properties of the Toll-like receptor adaptor protein TIRAP.** *J Biol Chem* 2009, **284**:9892-9898.
19. Snyder GA, Deredge D, Waldhuber A, Fresquez T, Wilkins DZ, Smith PT, Durr S, Cirl C, Jiang J, Jennings W et al.: **Crystal structures of the Toll/Interleukin-1 receptor (TIR) domains**

- from the *Brucella* protein TcpB and host adaptor TIRAP reveal mechanisms of molecular mimicry. *J Biol Chem* 2014, **289**:669-679.
20. Harte MT, Haga IR, Maloney G, Gray P, Reading PC, Bartlett NW, Smith GL, Bowie A, O'Neill LA: **The poxvirus protein A52R targets Toll-like receptor signaling complexes to suppress host defense.** *J Exp Med* 2003, **197**:343-351.
 21. Clark K, Nanda S, Cohen P: **Molecular control of the NEMO family of ubiquitin-binding proteins.** *Nat Rev Mol Cell Biol* 2013, **14**:673-685.
 22. Sen J, Liu X, Roller R, Knipe DM: **Herpes simplex virus US3 tegument protein inhibits Toll-like receptor 2 signaling at or before TRAF6 ubiquitination.** *Virology* 2013, **439**:65-73.
 23. Saito S, Murata T, Kanda T, Isomura H, Narita Y, Sugimoto A, Kawashima D, Tsurumi T: **Epstein-Barr virus deubiquitinase downregulates TRAF6-mediated NF-kappaB signaling during productive replication.** *J Virol* 2013, **87**:4060-4070.
 24. van Gent M, Braem SG, de Jong A, Delagic N, Peeters JG, Boer IG, Moynagh PN, Kremmer E, Wiertz EJ, Ovaas H *et al.*: **Epstein-Barr virus large tegument protein BPLF1 contributes to innate immune evasion through interference with Toll-like receptor signaling.** *PLoS Pathog* 2014, **10**:e1003960.
- Deubiquitinase activity of the N-terminus of BPLF1 was shown to remove K-48 and K-63 linked ubiquitin chains from TRAF6, NEMO, I κ B α blocking NF- κ B activation.
25. Sanada T, Kim M, Mimuro H, Suzuki M, Ogawa M, Oyama A, Ashida H, Kobayashi T, Koyama T, Nagai S *et al.*: **The *Shigella flexneri* effector OspI deamidates UBC13 to dampen the inflammatory response.** *Nature* 2012, **483**:623-626.
- OspI deamidates an active site glutamine of the E2 ubiquitin conjugase, UBC13, to glutamic acid. This novel mechanism prevents TRAF6 ubiquitination and blocks NF- κ B activation.
26. Zhang L, Ding X, Cui J, Xu H, Chen J, Gong YN, Hu L, Zhou Y, Ge J, Lu Q *et al.*: **Cysteine methylation disrupts ubiquitin-chain sensing in NF-kappaB activation.** *Nature* 2012, **481**:204-208.
- The cysteine methyl-transferase, NleE, of EPEC methylates zinc coordinating cysteines required for TAB2 and TAB3 to bind ubiquitin chains, thus preventing NF- κ B activation. This is the first description of enzymatic regulation of signal transduction via cysteine methylation.
27. Paquette N, Conlon J, Sweet C, Rus F, Wilson L, Pereira A, Rosadini CV, Goutagny N, Weber AN, Lane WS *et al.*: **Serine/threonine acetylation of TGFbeta-activated kinase (TAK1) by *Yersinia pestis* YopJ inhibits innate immune signaling.** *Proc Natl Acad Sci U S A* 2012, **109**:12710-12715.
- YopJ was found to acetylate both the *Drosophila* and mammalian MAP3K, TAK1. Acetylation of TAK1 at critical residues blocked IMD signaling in *Drosophila* and also prevented activation of mammalian TAK1. This work also demonstrated that YopJ targets multiple modules of signaling by not only blocking TAK1 but also acetylating the MAP2Ks, MKK4 and MKK7, effectively preventing activation of JNK, p38, and NF- κ B.
28. Reddick LE, Alto NM: **Bacteria fighting back: how pathogens target and subvert the host innate immune system.** *Mol Cell* 2014, **54**:321-328.
 29. Halfmann P, Neumann G, Kawaoka Y: **The Ebola virus VP24 protein blocks phosphorylation of p38 mitogen-activated protein kinase.** *J Infect Dis* 2011, **204**(Suppl. 3):S953-S956.
 30. Wang S, Chen Z, Hu C, Qian F, Cheng Y, Wu M, Shi B, Chen J, Hu Y, Yuan Z: **Hepatitis B virus surface antigen selectively inhibits TLR2 ligand-induced IL-12 production in monocytes/macrophages by interfering with JNK activation.** *J Immunol* 2013, **190**:5142-5151.
- Treatment of monocytes and macrophages with HBsAg inhibited Pam3csk4 stimulated JNK phosphorylation. JNK phosphorylation and IL-12 levels were also found to be reduced in patients with Hepatitis B infections. This is notable in that it suggests that viruses can interfere with TLR signaling at the MAPK signaling module.
31. Goncalves A, Burckstummer T, Dixit E, Scheicher R, Gorna MW, Karayel E, Sugar C, Stukalov A, Berg T, Kralovics R *et al.*: **Functional dissection of the TBK1 molecular network.** *PLoS ONE* 2011, **6**:e23971.
 32. Wang D, Fang L, Li K, Zhong H, Fan J, Ouyang C, Zhang H, Duan E, Luo R, Zhang Z *et al.*: **Foot-and-mouth disease virus 3C protease cleaves NEMO to impair innate immune signaling.** *J Virol* 2012, **86**:9311-9322.
 33. Wang D, Fang L, Wei D, Zhang H, Luo R, Chen H, Li K, Xiao S: **Hepatitis A virus 3C protease cleaves NEMO to impair induction of beta interferon.** *J Virol* 2014, **88**:10252-10258.
 34. Yamaguchi M, Kitagawa Y, Zhou M, Itoh M, Gotoh B: **An anti-interferon activity shared by paramyxovirus C proteins: inhibition of Toll-like receptor 7/9-dependent alpha interferon induction.** *FEBS Lett* 2014, **588**:28-34.
 35. Le Negrato G: **Viral interference with innate immunity by preventing NF-kappaB activity.** *Cell Microbiol* 2012, **14**:168-181.
 36. Taylor KE, Mossman KL: **Recent advances in understanding viral evasion of type I interferon.** *Immunology* 2013, **138**:190-197.
 37. Kim JC, Lee SY, Kim SY, Kim JK, Kim HJ, Lee HM, Choi MS, Min JS, Kim MJ, Choi HS *et al.*: **HSV-1 ICP27 suppresses NF-kappaB activity by stabilizing I κ B α .** *FEBS Lett* 2008, **582**:2371-2376.
 38. Graff JW, Ettayebi K, Hardy ME: **Rotavirus NSP1 inhibits NF-kappaB activation by inducing proteasome-dependent degradation of beta-TrCP: a novel mechanism of IFN antagonism.** *PLoS Pathog* 2009, **5**:e1000280.
 39. Mansur DS, Maluquer de Motes C, Unterholzner L, Sumner RP, Ferguson BJ, Ren H, Strnadova P, Bowie AG, Smith GL: **Poxvirus targeting of E3 ligase beta-TrCP by molecular mimicry: a mechanism to inhibit NF-kappaB activation and promote immune evasion and virulence.** *PLoS Pathog* 2013, **9**:e1003183.
 40. Vandevienne P, Lebrun M, El Mijiyad N, Ote I, Di Valentin E, Habraken Y, Dortu E, Piette J, Sadzot-Delvaux C: **The varicella-zoster virus ORF47 kinase interferes with host innate immune response by inhibiting the activation of IRF3.** *PLoS ONE* 2011, **6**:e16870.
 41. Wang S, Wang K, Lin R, Zheng C: **Herpes simplex virus 1 serine/threonine kinase US3 hyperphosphorylates IRF3 and inhibits beta interferon production.** *J Virol* 2013, **87**:12814-12827.
 42. Wang K, Ni L, Wang S, Zheng C: **Herpes simplex virus 1 protein kinase US3 hyperphosphorylates p65/RelA and dampens NF-kappaB activation.** *J Virol* 2014, **88**:7941-7951.
 43. Melroe GT, Silva L, Schaffer PA, Knipe DM: **Recruitment of activated IRF-3 and CBP/p300 to herpes simplex virus ICPO nuclear foci: potential role in blocking IFN-beta induction.** *Virology* 2007, **360**:305-321.
 44. Lin R, Genin P, Mamane Y, Sgarbanti M, Battistini A, Harrington WJ Jr, Barber GN, Hiscott J: **HHV-8 encoded vIRF-1 represses the interferon antiviral response by blocking IRF-3 recruitment of the CBP/p300 coactivators.** *Oncogene* 2001, **20**:800-811.
 45. Xing J, Ni L, Wang S, Wang K, Lin R, Zheng C: **Herpes simplex virus 1-encoded tegument protein VP16 abrogates the production of beta interferon (IFN) by inhibiting NF-kappaB activation and blocking IFN regulatory factor 3 to recruit its coactivator CBP.** *J Virol* 2013, **87**:9788-9801.
 46. Gao X, Wan F, Mateo K, Callegari E, Wang D, Deng W, Puente J, Li F, Chaussee MS, Finlay BB *et al.*: **Bacterial effector binding to ribosomal protein s3 subverts NF-kappaB function.** *PLoS Pathog* 2009, **5**:e1000708.
 47. Royan SV, Jones RM, Koutsouris A, Roxas JL, Falzari K, Weflen AW, Kim A, Bellmeyer A, Turner JR, Neish AS *et al.*: **Enteropathogenic *E. coli* non-LEE encoded effectors NleH1 and NleH2 attenuate NF-kappaB activation.** *Mol Microbiol* 2010, **78**:1232-1245.
 48. Muhlen S, Ruchaud-Sparagano MH, Kenny B: **Proteasome-independent degradation of canonical NF-kappaB complex components by the NleC protein of pathogenic *Escherichia coli*.** *J Biol Chem* 2011, **286**:5100-5107.
 49. Pearson JS, Riedmaier P, Marches O, Frankel G, Hartland EL: **A type III effector protease NleC from enteropathogenic**

- Escherichia coli* targets NF-kappaB for degradation.** *Mol Microbiol* 2011, **80**:219-230.
50. Yen H, Ooka T, Iguchi A, Hayashi T, Sugimoto N, Tobe T: **NleC, a type III secretion protease, compromises NF-kappaB activation by targeting p65/RelA.** *PLoS Pathog* 2010, **6**:e1001231.
 51. Baruch K, Gur-Arie L, Nadler C, Koby S, Yerushalmi G, Ben-Neriah Y, Yogeve O, Shaulian E, Guttman C, Zarivach R *et al.*: **Metalloprotease type III effectors that specifically cleave JNK and NF-kappaB.** *EMBO J* 2011, **30**:221-231.
 52. Newton HJ, Pearson JS, Badea L, Kelly M, Lucas M, Holloway G, Wagstaff KM, Dunstone MA, Sloan J, Whisstock JC *et al.*: **The type III effectors NleE and NleB from enteropathogenic *E. coli* and OspZ from *Shigella* block nuclear translocation of NF-kappaB p65.** *PLoS Pathog* 2010, **6**:e1000898.
 53. Kim DW, Lenzen G, Page AL, Legrain P, Sansonetti PJ, Parsot C: **The *Shigella flexneri* effector OspG interferes with innate immune responses by targeting ubiquitin-conjugating enzymes.** *Proc Natl Acad Sci U S A* 2005, **102**:14046-14051.
 54. Pruneda JN, Smith FD, Daurie A, Swaney DL, Villen J, Scott JD, Stadnyk AW, Le Trong I, Stenkamp RE, Klevit RE *et al.*: **E2~Ub conjugates regulate the kinase activity of *Shigella* effector OspG during pathogenesis.** *EMBO J* 2014, **33**:437-449.
 55. DiPerna G, Stack J, Bowie AG, Boyd A, Kotwal G, Zhang Z, Arvikar S, Latz E, Fitzgerald KA, Marshall WL: **Poxvirus protein N1L targets the I-kappaB kinase complex, inhibits signaling to NF-kappaB by the tumor necrosis factor superfamily of receptors, and inhibits NF-kappaB and IRF3 signaling by toll-like receptors.** *J Biol Chem* 2004, **279**:36570-36578.
 56. Newman RM, Salunkhe P, Godzik A, Reed JC: **Identification and characterization of a novel bacterial virulence factor that shares homology with mammalian Toll/interleukin-1 receptor family proteins.** *Infect Immun* 2006, **74**:594-601.
 57. Li H, Xu H, Zhou Y, Zhang J, Long C, Li S, Chen S, Zhou JM, Shao F: **The phosphothreonine lyase activity of a bacterial type III effector family.** *Science* 2007, **315**:1000-1003.
 58. Mazurkiewicz P, Thomas J, Thompson JA, Liu M, Arbibe L, Sansonetti P, Holden DW: **SpvC is a *Salmonella* effector with phosphothreonine lyase activity on host mitogen-activated protein kinases.** *Mol Microbiol* 2008, **67**:1371-1383.
 59. Mukherjee S, Keitany G, Li Y, Wang Y, Ball HL, Goldsmith EJ, Orth K: ***Yersinia* YopJ acetylates and inhibits kinase activation by blocking phosphorylation.** *Science* 2006, **312**:1211-1214.
 60. Tobe T, Beatson SA, Taniguchi H, Abe H, Bailey CM, Fivian A, Younis R, Matthews S, Marches O, Frankel G *et al.*: **An extensive repertoire of type III secretion effectors in *Escherichia coli* O157 and the role of lambdoid phages in their dissemination.** *Proc Natl Acad Sci U S A* 2006, **103**:14941-14946.
 61. Matsumoto H, Young GM: **Translocated effectors of *Yersinia*.** *Curr Opin Microbiol* 2009, **12**:94-100.
 62. Datta A, Sinha-Datta U, Dhillon NK, Buch S, Nicot C: **The HTLV-I p30 interferes with TLR4 signaling and modulates the release of pro- and anti-inflammatory cytokines from human macrophages.** *J Biol Chem* 2006, **281**:23414-23424.
 63. Peri P, Mattila RK, Kantola H, Broberg E, Karttunen HS, Waris M, Vuorinen T, Hukkanen V: **Herpes simplex virus type 1 Us3 gene deletion influences toll-like receptor responses in cultured monocytic cells.** *Virology* 2008, **5**:140.
 64. Yokoyama R, Itoh S, Kamoshida G, Takii T, Fujii S, Tsuji T, Onozaki K: **Staphylococcal superantigen-like protein 3 binds to the Toll-like receptor 2 extracellular domain and inhibits cytokine production induced by *Staphylococcus aureus*, cell wall component, or lipopeptides in murine macrophages.** *Infect Immun* 2012, **80**:2816-2825.
- This paper demonstrates that purified SSL3 of *S. aureus* binds to the ectodomain of TLR2 and blocks signaling in leukocytes. These works represent the first example of a bacterial protein that can inhibit TLR signaling by antagonizing the receptor module.
65. Bardoel BW, Vos R, Bouman T, Aerts PC, Bestebroer J, Huizinga EG, Brondijk TH, van Strijp JA, de Haas CJ: **Evasion of Toll-like receptor 2 activation by staphylococcal superantigen-like protein 3.** *J Mol Med (Berl)* 2012, **90**:1109-1120.
- See annotation to Ref. [63**].
66. Bestebroer J, van Kessel KP, Azouagh H, Walenkamp AM, Boer IG, Romijn RA, van Strijp JA, de Haas CJ: **Staphylococcal SSL5 inhibits leukocyte activation by chemokines and anaphylatoxins.** *Blood* 2009, **113**:328-337.
 67. Kilian M, Reinholdt J, Lomholt H, Poulsen K, Frandsen EV: **Biological significance of IgA1 proteases in bacterial colonization and pathogenesis: critical evaluation of experimental evidence.** *APMIS* 1996, **104**:321-338.
 68. Clementi CF, Murphy TF: **Non-typeable *Haemophilus influenzae* invasion and persistence in the human respiratory tract.** *Front Cell Infect Microbiol* 2011, **1**:1.
 69. Wilensky A, Tzach-Nahman R, Potempa J, Shapira L, Nussbaum G: ***Porphyromonas gingivalis* gingipains selectively reduce CD14 expression, leading to macrophage hyporesponsiveness to bacterial infection.** *J Innate Immun* 2014.
 70. de Oliveira VL, Almeida SC, Soares HR, Crespo A, Marshall-Clarke S, Parkhouse RM: **A novel TLR3 inhibitor encoded by African swine fever virus (ASFV).** *Arch Virol* 2011, **156**:597-609.
 71. Xiang Z, Li L, Lei X, Zhou H, Zhou Z, He B, Wang J: **Enterovirus 68 3C protease cleaves TRIF to attenuate antiviral responses mediated by Toll-like receptor 3.** *J Virol* 2014, **88**:6650-6659.
 72. Li K, Foy E, Ferreone JC, Nakamura M, Ferreone AC, Ikeda M, Ray SC, Gale M Jr, Lemon SM: **Immune evasion by hepatitis C virus NS3/4A protease-mediated cleavage of the Toll-like receptor 3 adaptor protein TRIF.** *Proc Natl Acad Sci U S A* 2005, **102**:2992-2997.
 73. Abe T, Kaname Y, Hamamoto I, Tsuda Y, Wen X, Tagawa S, Moriishi K, Takeuchi O, Kawai T, Kanto T *et al.*: **Hepatitis C virus nonstructural protein 5A modulates the toll-like receptor-MYD88-dependent signaling pathway in macrophage cell lines.** *J Virol* 2007, **81**:8953-8966.
 74. Ahmad H, Gubbels R, Ehlers E, Meyer F, Waterbury T, Lin R, Zhang L: **Kaposi sarcoma-associated herpesvirus degrades cellular Toll-interleukin-1 receptor domain-containing adaptor-inducing beta-interferon (TRIF).** *J Biol Chem* 2011, **286**:7865-7872.
 75. Salcedo SP, Marchesini MI, Degos C, Terwagne M, Von Bargen K, Lepidi H, Herrmann CK, Santos Lacerda TL, Imbert PR, Pierre P *et al.*: **BtpA, a novel *Brucella* TIR-containing effector protein with immune modulatory functions.** *Front Cell Infect Microbiol* 2013, **3**:28.
 76. Cirl C, Wieser A, Yadav M, Duerr S, Schubert S, Fischer H, Stappert D, Wantia N, Rodriguez N, Wagner H *et al.*: **Subversion of Toll-like receptor signaling by a unique family of bacterial Toll/interleukin-1 receptor domain-containing proteins.** *Nat Med* 2008, **14**:399-406.
 77. Duesbery NS, Webb CP, Leppla SH, Gordon VM, Klimpel KR, Copeland TD, Ahn NG, Oskarsson MK, Fukasawa K, Paull KD *et al.*: **Proteolytic inactivation of MAP-kinase-kinase by anthrax lethal factor.** *Science* 1998, **280**:734-737.
 78. Jones RM, Wu H, Wentworth C, Luo L, Collier-Hyams L, Neish AS: ***Salmonella* AvrA coordinates suppression of host immune and apoptotic defenses via JNK pathway blockade.** *Cell Host Microbe* 2008, **3**:233-244.
 79. Arbibe L, Kim DW, Batsche E, Pedron T, Mateescu B, Muchardt C, Parsot C, Sansonetti PJ: **An injected bacterial effector targets chromatin access for transcription factor NF-kappaB to alter transcription of host genes involved in immune responses.** *Nat Immunol* 2007, **8**:47-56.
 80. Trosky JE, Li Y, Mukherjee S, Keitany G, Ball H, Orth K: **VopA inhibits ATP binding by acetylating the catalytic loop of MAPK kinases.** *J Biol Chem* 2007, **282**:34299-34305.
 81. Otsuka M, Kato N, Moriyama M, Taniguchi H, Wang Y, Dharel N, Kawabe T, Omata M: **Interaction between the HCV NS3 protein**

- and the host TBK1 protein leads to inhibition of cellular antiviral responses. *Hepatology* 2005, **41**:1004-1012.
82. Ma Y, Jin H, Valyi-Nagy T, Cao Y, Yan Z, He B: **Inhibition of TANK binding kinase 1 by herpes simplex virus 1 facilitates productive infection.** *J Virol* 2012, **86**:2188-2196.
 83. Wang G, Chen G, Zheng D, Cheng G, Tang H: **PLP2 of mouse hepatitis virus A59 (MHV-A59) targets TBK1 to negatively regulate cellular type I interferon signaling pathway.** *PLoS ONE* 2011, **6**:e17192.
 84. Chen RA, Ryzhakov G, Cooray S, Randow F, Smith GL: **Inhibition of I kappa B kinase by vaccinia virus virulence factor B14.** *PLoS Pathog* 2008, **4**:e22.
 85. Unterholzner L, Sumner RP, Baran M, Ren H, Mansur DS, Bourke NM, Randow F, Smith GL, Bowie AG: **Vaccinia virus protein C6 is a virulence factor that binds TBK-1 adaptor proteins and inhibits activation of IRF3 and IRF7.** *PLoS Pathog* 2011, **7**:e1002247.
 86. Schroder M, Baran M, Bowie AG: **Viral targeting of DEAD box protein 3 reveals its role in TBK1/IKKepsilon-mediated IRF activation.** *EMBO J* 2008, **27**:2147-2157.
 87. Ashida H, Kim M, Schmidt-Supprian M, Ma A, Ogawa M, Sasakawa C: **A bacterial E3 ubiquitin ligase IpaH9.8 targets NEMO/IKKgamma to dampen the host NF-kappaB-mediated inflammatory response.** *Nat Cell Biol* 2010, **12**:66-73 sup 99 61-69.
 88. Mittal R, Peak-Chew SY, McMahon HT: **Acetylation of MEK2 and I kappa B kinase (IKK) activation loop residues by YopJ inhibits signaling.** *Proc Natl Acad Sci U S A* 2006, **103**:18574-18579.
 89. Bauhofer O, Summerfield A, Sakoda Y, Tratschin JD, Hofmann MA, Ruggli N: **Classical swine fever virus Npro interacts with interferon regulatory factor 3 and induces its proteasomal degradation.** *J Virol* 2007, **81**:3087-3096.
 90. Okumura A, Alce T, Lubyova B, Ezelle H, Strebel K, Pitha PM: **HIV-1 accessory proteins VPR and Vif modulate antiviral response by targeting IRF-3 for degradation.** *Virology* 2008, **373**:85-97.
 91. Doehle BP, Chang K, Rustagi A, McNeven J, McElrath MJ, Gale M Jr: **Vpu mediates depletion of interferon regulatory factor 3 during HIV infection by a lysosome-dependent mechanism.** *J Virol* 2012, **86**:8367-8374.
 92. Hotter D, Kirchhoff F, Sauter D: **HIV-1 Vpu does not degrade interferon regulatory factor 3.** *J Virol* 2013, **87**:7160-7165.
 93. Yu Y, Hayward GS: **The ubiquitin E3 ligase RAUL negatively regulates type I interferon through ubiquitination of the transcription factors IRF7 and IRF3.** *Immunity* 2010, **33**:863-877.
 94. Lefort S, Soucy-Faulkner A, Grandvaux N, Flamand L: **Binding of Kaposi's sarcoma-associated herpesvirus K-bZIP to interferon-responsive factor 3 elements modulates antiviral gene expression.** *J Virol* 2007, **81**:10950-10960.
 95. Joo CH, Shin YC, Gack M, Wu L, Levy D, Jung JU: **Inhibition of interferon regulatory factor 7 (IRF7)-mediated interferon signal transduction by the Kaposi's sarcoma-associated herpesvirus viral IRF homolog vIRF3.** *J Virol* 2007, **81**:8282-8292.
 96. Sparrer KM, Pfaller CK, Conzelmann KK: **Measles virus C protein interferes with Beta interferon transcription in the nucleus.** *J Virol* 2012, **86**:796-805.
 97. Irie T, Kiyotani K, Igarashi T, Yoshida A, Sakaguchi T: **Inhibition of interferon regulatory factor 3 activation by paramyxovirus V protein.** *J Virol* 2012, **86**:7136-7145.
 98. Lu LL, Puri M, Horvath CM, Sen GC: **Select paramyxoviral V proteins inhibit IRF3 activation by acting as alternative substrates for inhibitor of kappaB kinase epsilon (IKKe)/TBK1.** *J Biol Chem* 2008, **283**:14269-14276.
 99. Barro M, Patton JT: **Rotavirus NSP1 inhibits expression of type I interferon by antagonizing the function of interferon regulatory factors IRF3, IRF5, and IRF7.** *J Virol* 2007, **81**:4473-4481.
 100. Arnold MM, Patton JT: **Diversity of interferon antagonist activities mediated by NSP1 proteins of different rotavirus strains.** *J Virol* 2011, **85**:1970-1979.
 101. Zhu H, Zheng C, Xing J, Wang S, Li S, Lin R, Mossman KL: **Varicella-zoster virus immediate-early protein ORF61 abrogates the IRF3-mediated innate immune response through degradation of activated IRF3.** *J Virol* 2011, **85**:11079-11089.
 102. Sloan E, Henriquez R, Kinchington PR, Slobedman B, Abendroth A: **Varicella-zoster virus inhibition of the NF-kappaB pathway during infection of human dendritic cells: role for open reading frame 61 as a modulator of NF-kappaB activity.** *J Virol* 2012, **86**:1193-1202.